

and  $\Delta H_3$ ; thus the hydrogen bond formed between a phenol and a homoconjugated anion is very weak.

The heat evolved in the formation of the first hydrogen bond between a phenolate ion and a phenol  $\Delta H_4$  is significantly larger than that of the proton transfer from the phenol to the acridine orange base (Table 2). Therefore, the energetics of reaction (2) are mainly determined by this association. The association tendency of the phenolate ions in benzonitrile is much stronger than in acetonitrile. This is due to a larger reaction enthalpy  $\Delta H_4$ . The difference of about  $8 \text{ kJ mol}^{-1}$  between  $\Delta H_4$  in these solvents may be ascribed to different hydrogen bond energies of hydrogen bonds between the phenol monomers and the solvents since benzonitrile is a weaker hydrogen bond acceptor than acetonitrile.

The solvent dependence of the *AO/NP* system reveals a surprising effect of the homoconjugation. At the ionic strength 0.1 M,  $K_1$  has almost the same value in both solvents. This is a fortuitous coincidence, since the dependence of  $K_1$  on the ionic strength in these solvents is quite different. The concentration of ions in a benzonitrile solution of *AO* and *NP* at the ionic strength 0.1 M is significantly larger than in a solution with the same starting concentrations in acetonitrile. This is due solely to the stronger anion association which shifts the overall equilibrium to the side of the ions in spite of the smaller dielectric constant of benzonitrile.

The authors are indebted to Prof. H. W. Zimmermann for his constant encouragement. Financial support by the Deutsche Forschungsgemeinschaft, the Fonds der Chemischen Industrie and the Freiburger Wissenschaftliche Gesellschaft is gratefully acknowledged. R. S. gratefully acknowledges a stipend of the Studienstiftung des Deutschen Volkes. The calculations were carried out at the Rechenzentrum of the Universität Freiburg.

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(Eingegangen am 15. August 1983, E 5540  
endgültige Fassung am 2. Mai 1984)

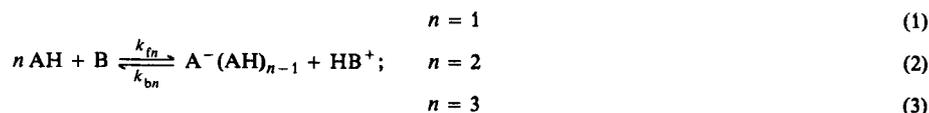
## Acid Base Reactions Between Acridine Orange and Substituted Phenols in Benzonitrile Part II. Kinetics: An Extended Eigen Mechanism of Proton Transfer

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### *Chemical Kinetics / Elementary Reactions / Molecular Interactions / Solutions*

The kinetics of proton transfer between five substituted phenols (AH) and the acridine orange base (B) are studied in benzonitrile solution using the temperature jump method. In the overall reactions (1) – (3)



the formation and dissociation of an intermediate ion pair  $A^- \cdot HB^+$  is rate determining. The rate constants of the bimolecular recombination of the ions to the ion pair are  $k_{b1}$ ,  $k_{b2}$ , and  $k_{b3}$ . Their values are in the range  $(0.18 - 1.7) \cdot 10^9 \text{ M}^{-1} \text{ s}^{-1}$  ( $25^\circ\text{C}$ ,  $I = 0.1 \text{ M}$ ).  $k_{b2}$  and  $k_{b3}$  do not depend on the  $pK$  of the phenol involved, although these rates are not diffusion controlled. The rate of proton transfer from the protonated acridine orange ( $BH^+$ ) to the free anions ( $A^-$ ) does depend to some degree on the acidity of the corresponding phenol. In the case of the 3-methyl-4-nitrophenolate ion this rate is diffusion controlled, as shown by the dependence of  $k_{b1}$  on the temperature and the ionic strength. Lower limits for the formation constants of the ion pairs are estimated from the rate data. The equilibria and the kinetics are discussed in terms of an extended Eigen scheme of proton transfer, and the separate steps of anion solvation by hydrogen bonding to phenol molecules shown to depend in different ways on the basicity of the anion.

### 1. Introduction

Proton transfer reactions between organic acids and bases in aqueous solutions were among the first classes of chemical reactions to be extensively studied by relaxation methods [1, 2]. Diffusion controlled rates are a general feature of these reactions [3]. In organic solvents different mechanisms of proton transfer are observed. The rate constants of ion pair formation of many amines with phenols in halogenated hydrocarbon solvents are ten to hundred times less than the calculated values for diffusion control, and do not correlate with the equilibrium constants [4, 5]. This is explained by diffusion controlled formation of weakly bound intermediate complexes with sufficient lifetime to allow rotation of the reactants within the complex [5]. Recently we have found that proton transfers between protonated nitrogen bases and phenolate ions in dry acetonitrile are kinetically hindered by the need for solvent reorientation at the site of the proton, whereas there is no measurable energy barrier for the transfer of the proton within the hydrogen bonded complex [6, 7].

Similar effects could be expected to be operative in benzonitrile solution, since the dipole moment of the benzonitrile molecule is almost equal to that of the acetonitrile molecule and since molecular reorientation is slower in benzonitrile because of its larger viscosity. A strong solvation of ions by benzonitrile is indicated by the solubility of many salts in this solvent, in spite of its relatively low dielectric constant ( $\epsilon = 25.2$  at  $25^\circ\text{C}$ ). No investigation of the kinetics of proton transfer reactions in benzonitrile has previously been reported.

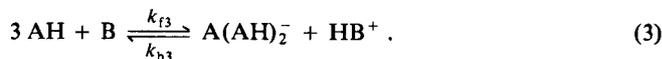
The following is a report of the conclusions of a study of the kinetics and energetics of proton transfers between 3-chloro-4-nitrophenol *CNP*, 4-nitrophenol *NP*, 3-methyl-4-nitrophenol *MNP*, 4-chloro-3,5-dimethylphenol *CMP*, and 3,5-dimethylphenol *DMP* and the indicator base acridine orange in benzonitrile. Depending on the concentration of the phenol  $AH$  proton transfer is observed in three different overall reactions [8]. At concentrations  $c_{AH} < 10^{-3} \text{ M}$  free ions are formed by protonation of the base  $B$



together with the homoconjugate anions  $AHA^-$



At higher concentrations of the phenol reaction (1) could not be observed; however, in addition to reaction (2), further association of the anions occurred according to



These reactions are all exothermic. Some of them are nevertheless endergonic because of the large negative entropy connected with ion formation in benzonitrile. The equilibrium- and thermodynamic data of the acid base and association reactions have been given in the preceding paper [8].

### 2. Experimental

Purification of the solvent and of the other chemicals has been described [8]. The kinetic measurements were done with a Meßanlagen temperature jump transient spectrometer. The ionic strength of the sample solutions was kept constant by adding  $Bu_4NPF_6$  *TBHP*. The viscosities of solutions of *TBHP* in benzonitrile were measured at  $25^\circ\text{C}$

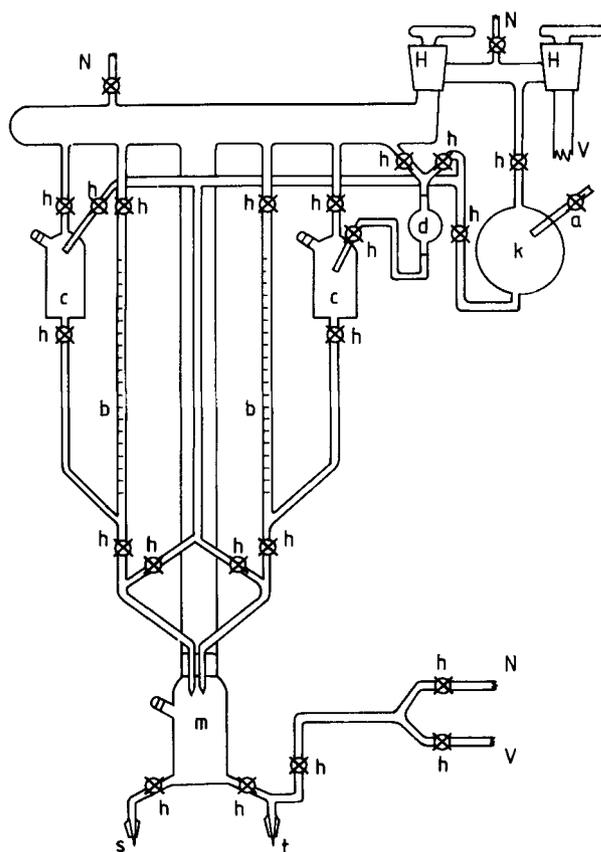


Fig. 1

Buret system for preparing sample solutions in an inert atmosphere. The freshly distilled benzonitrile is filled into the storage bulb  $k$  through valve  $a$  by aid of a tefton tubing;  $b$  = burets;  $c$  = stock flask for preparing reagent solutions;  $d$  = calibrated flask that allows filling a measured volume of solvent into  $c$ ;  $h$  = tefton valve;  $H$  = 15 mm glass valve;  $m$  = mixing chamber with outlets  $s$  to spectrophotometric cell and  $t$  to  $T$ -jump cell;  $N$  = nitrogen inlets;  $V$  = connections to vacuum line

with an Ubbelohde viscosimeter. Contamination of the sample solutions with moisture was avoided by preparing stocks and the samples in the closed all glass apparatus shown in Fig. 1. This apparatus was dried before each preparation by evacuating it for at least 12 hours at about  $10^{-2}$  Pa. In each sample solution the equilibrium concentration of  $\text{HB}^+$  was checked spectrophotometrically. The solution was not used to evaluate kinetic data when the equilibrium constant calculated from this measurement deviated by more than  $\pm 10\%$  from the value previously obtained [8].

The  $T$ -jump cell was thermostatted by circulating water through the upper electrode. The temperature before the jump was measured by means of a Si-diode sealed in a teflon tubing dipping into the solution near to the upper electrode. Charging voltages of 30 kV and 35 kV were used, resulting in respective temperature increases of  $3.2^\circ\text{C}$  and  $4.4^\circ\text{C}$ . The signals were stored in a Biomation transient recorder Model 805, and evaluated with a non-linear least squares program, which allows data fitting with one or two exponentials. 800 to 1200 data points were taken for each signal. In calculating the relaxation times  $\tau_2$  the heating process was taken into account as the first exponential ( $\tau_1$ ). The heating times were calculated from the measured resistance as [9]

$$\tau_e = 1/2 RC.$$

In some instances both the relaxation times  $\tau_1$  and  $\tau_2$  were calculated by curve fitting.  $\tau_1$  obtained in this way agreed to within a few percent with  $\tau_e$ . A typical signal is shown in Fig. 2. Between three and six signals were taken for each sample solution. The relaxation times for a particular sample solution agreed typically better than  $\pm 5\%$  of the average relaxation time.

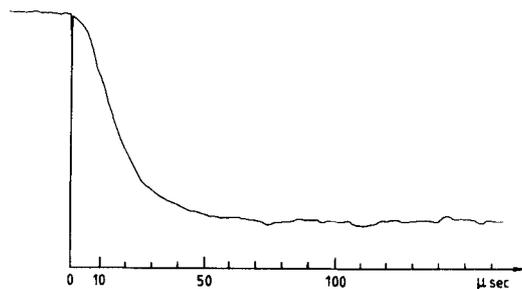


Fig. 2  
Relaxation curve for the proton transfers between AO and NP,  
 $\tau = 10.9 \mu\text{sec}$

### 3. Results

#### 3.1. Proton Transfer Between Acridine Orange and Nitrophenols

For the protonation of the acridine orange base B by *CNP*, *NP* and *MNP* concentrations of  $10^{-5}$ – $10^{-4}$  M of both reactants were used. The reaction was monitored with the absorption of the protonated acridine orange  $\text{BH}^+$  at  $20000 \text{ cm}^{-1}$ . In all solutions only one chemical relaxation signal was observed. It is ascribed to the parallel reactions (1) and (2). These are coupled by the hydrogen bond association of the phenolate anion  $\text{A}^-$  to the homoconjugate ion  $\text{AHA}^-$ :



This reaction was studied independently. Solutions containing *NP* and its tetrabutylammoniumsalt show a relaxation time after a temperature jump which is very close to the heating time.

Therefore only a lower limit for  $k_{f4}$  could be determined;  $k_{f4}^{\text{NP}}$  is greater than  $4 \cdot 10^8 \text{ M}^{-1} \text{ s}^{-1}$ . Under the conditions used to study the acid base reactions (1) and (2) the equilibration of (4) is rapid compared with that of (1) and (2). The reciprocal relaxation time,  $1/\tau$  for reac-

tions (1) and (2) is calculated by standard methods of relaxation kinetics [1]

$$1/\tau = k_0 F_1 \quad (6)$$

In (6) the concentration term

$$F_1 = K_1 \cdot \left( c_{\text{AH}} + \frac{c_{\text{B}}(2c_{\text{AH}} + K_4^{-1})}{c_{\text{AH}} + c_{\text{A}^-} + K_4^{-1}} \right) + c_{\text{A}^-} + \frac{c_{\text{HB}} + (2c_{\text{A}^-} + K_4^{-1})}{c_{\text{AH}} + c_{\text{A}^-} + K_4^{-1}}$$

accounts for the preequilibrium, and the observed rate constant  $k_0$  is defined by

$$k_0 = k_{b1} + k_{b2} \cdot K_4 \cdot c_{\text{AH}}.$$

Values of  $k_{b1}$  and  $k_{b2}$  are obtained from intercept and slope of linear plots of Eq. (7)

$$(\tau \cdot F_1)^{-1} = k_{b1} + k_{b2} \cdot K_4 \cdot c_{\text{AH}} \quad (7)$$

shown in Fig. 3. The equilibrium relations  $k_{f1} = K_1 \cdot k_{b1}$  and  $k_{f2} = K_2 \cdot k_{b2}$  can be used to calculate  $k_{f1}$  and  $k_{f2}$ . The results are given in Table 2. The  $\tau$  values are listed in Table 1. The agreement between experimental and calculated values is very good.

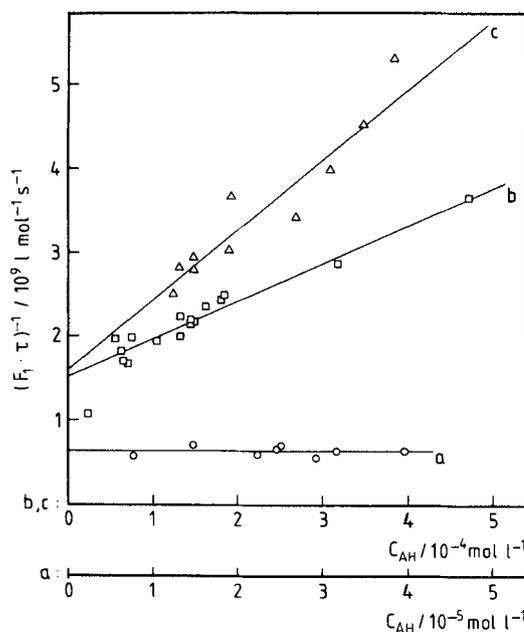


Fig. 3  
Dependence of the relaxation times of proton transfers between nitrophenols and Acridine Orange on the phenol concentration. Plot of the rate expression Eq. (7) vs. the equilibrium concentration of a) *CNP*, b) *NP*, and c) *MNP*

In the experiments with *CNP* the concentration of the phenol had to be kept very low in order to avoid complete protonation of the base. Under these conditions reaction (2) did not make a measurable contribution to the observed relaxation. This is evident from Fig. 3, where the plot of Eq. (7) for the *CNP* system produces a zero slope. The standard errors of the estimated relaxation times are  $\sigma = 0.068$ ,  $0.066$ , and  $0.066$  for *CNP*, *NP* and *MNP* respectively.

The rates of proton transfer between *MNP* and acridine orange were measured also at  $18^\circ\text{C}$ ,  $35^\circ\text{C}$ , and  $50^\circ\text{C}$  (Fig. 4). Rate constants were calculated using Eqs. (6) and (7) and the equilibrium relations as before. In this temperature range we obtain (Fig. 5)

$$k_{b1} = (3.19 \pm 0.09) \cdot 10^{11} \exp(-12.8 \pm 1.8 \text{ kJ mol}^{-1}/RT) \text{ M}^{-1} \text{ s}^{-1}$$

and

$$k_{b2} = (2.82 \pm 0.08) \cdot 10^{15} \exp(-40.0 \pm 2.6 \text{ kJ mol}^{-1}/RT) \text{ M}^{-1} \text{ s}^{-1}.$$

Table I  
Reactions of Acridine Orange base with Nitrophenols

$10^4 c_{\text{AH}}^0$ <sup>a)</sup> M	$10^5 c_{\text{B}}^0$ <sup>b)</sup> M	$\tau$ $\mu\text{sec}$
3-Cl-4-nitrophenol, $T = 25^\circ\text{C}$ , $I = 0.1 \text{ M}$		
0.608	3.19	29.3
0.365	4.13	26.4
0.632	5.97	19.7
1.00	7.19	18.2
0.807	10.2	13.9
1.03	13.2	13.6
0.909	16.6	11.1
0.469	25.1	7.7
4-Nitrophenol, $T = 25^\circ\text{C}$ , $I = 0.1 \text{ M}$		
0.378	1.09	52.5
1.80	1.40	51.1
1.28	2.90	46.0
2.10	2.56	36.1
1.81	3.66	35.0
2.17	2.94	34.1
0.842	10.2	25.0
1.93	6.67	24.8
1.84	8.44	21.6
2.02	13.0	18.1
1.28	19.3	17.0
1.04	14.7	16.9
1.25	18.7	16.5
3.92	5.86	16.5
2.24	16.2	14.8
1.22	27.6	12.5
5.66	0.624	10.9
3-CH <sub>3</sub> -4-nitrophenol, $T = 25^\circ\text{C}$ , $I = 0.1 \text{ M}$		
2.86	1.45	72.6
1.67	2.92	63.3
3.86	3.17	36.5
3.72	1.36	44.7
2.48	10.61	30.4
1.98	14.6	27.4
3.64	6.94	27.4
1.74	21.3	25.3
2.23	14.7	25.1
4.53	5.65	24.1
1.83	18.9	23.3
6.75	3.88	20.9
3-CH <sub>3</sub> -4-nitrophenol, $T = 18^\circ\text{C}$ , $I = 0.1 \text{ M}$		
1.29	3.62	79.3
2.29	1.96	72.9
2.71	4.44	46.0
4.50	8.09	25.1
4.39	12.5	21.3

Table I (Continued)

$10^4 c_{\text{AH}}^0$ <sup>a)</sup> M	$10^5 c_{\text{B}}^0$ <sup>b)</sup> M	$\tau$ $\mu\text{sec}$
3-CH <sub>3</sub> -4-nitrophenol, $T = 35^\circ\text{C}$ , $I = 0.1 \text{ M}$		
1.21	4.33	50.7
2.45	3.50	42.2
3.54	5.84	26.2
5.62	6.38	18.1
4.61	4.28	24.8
4.61	8.77	17.9
6.59	8.48	14.4
7.09	12.1	12.0
6.16	15.6	11.2
3-CH <sub>3</sub> -4-nitrophenol, $T = 50^\circ\text{C}$ , $I = 0.1 \text{ M}$		
4.07	2.23	29.8
6.07	2.82	20.4
7.90	4.06	13.8
10.3	4.67	9.5
11.3	6.58	8.3
3-CH <sub>3</sub> -4-nitrophenol, $T = 25^\circ\text{C}$ , $I = 0.025 \text{ M}$		
1.74	4.46	42.5
2.32	2.81	39.4
3.00	2.21	38.0
3.69	5.47	20.0
4.50	4.19	19.2
5.48	5.80	16.5
3-CH <sub>3</sub> -4-nitrophenol, $T = 25^\circ\text{C}$ , $I = 0.05 \text{ M}$		
1.79	3.35	52.0
2.75	1.90	49.2
2.79	2.11	44.8
2.74	6.69	25.9
4.53	3.85	24.2
3.98	4.40	24.0
3.26	11.1	18.5
3.53	10.5	18.3
7.51	3.87	14.8
3-CH <sub>3</sub> -4-nitrophenol, $T = 25^\circ\text{C}$ , $I = 0.2 \text{ M}$		
1.03	4.46	82.7
1.75	2.73	77.6
2.66	4.42	44.5
1.77	9.78	44.2
2.10	7.50	43.0
2.27	11.7	31.4

a) Total concentration of phenol.

b) Total concentration of Acridine Orange.

Table II  
Rate and Equilibrium Constants for the Reactions of Acridine Orange with Nitrophenols in Benzonitrile<sup>a)</sup>

Phenol	$T$ $^\circ\text{C}$	$I$ M	$10^{-7} k_{f1}$ $\text{M}^{-1} \text{s}^{-1}$	$10^{-8} k_{b1}$ $\text{M}^{-1} \text{s}^{-1}$	$K_1 = k_{f1}/k_{b1}$ <sup>b)</sup>	$10^{-10} k_{f2}$ $\text{M}^{-2} \text{s}^{-1}$	$10^{-8} k_{b2}$ $\text{M}^{-1} \text{s}^{-1}$	$K_2 = k_{f2}/k_{b2}$ <sup>b)</sup> $\text{M}^{-1}$
CNP	25	0.1	$39.0 \pm 4$	$5.9 \pm 0.6$	0.66	—	—	15600
NP	25	0.1	$4.0 \pm 0.08$	$14.6 \pm 0.3$	0.0276	$13.3 \pm 0.3$	$1.9 \pm 0.07$	690
MNP	25	0.1	$1.1 \pm 0.05$	$16.9 \pm 0.6$	0.0064	$4.9 \pm 0.1$	$3.1 \pm 0.07$	155
	18	0.1	$1.4 \pm 0.06$	$16.6 \pm 0.8$	0.0084	$4.5 \pm 0.3$	$1.8 \pm 0.1$	253
	35	0.1	$1.3 \pm 0.03$	$24.4 \pm 0.5$	0.0055	$3.2 \pm 0.06$	$4.0 \pm 0.1$	80
	50	0.1	$1.0 \pm 0.1$	$26.4 \pm 2.6$	0.0040	$3.3 \pm 0.2$	$10.2 \pm 0.4$	32
	25	0.025	$1.2 \pm 0.1$	$39.7 \pm 3.9$	0.0030	$5.0 \pm 0.3$	$5.7 \pm 0.3$	88
	25	0.05	$1.0 \pm 0.02$	$25.1 \pm 0.5$	0.0040	$5.6 \pm 0.06$	$4.8 \pm 0.05$	117
	25	0.2	$1.0 \pm 0.05$	$8.7 \pm 0.5$	0.011	$6.5 \pm 0.3$	$2.5 \pm 0.15$	260

a) The errors are standard errors. b) From Ref. [8].

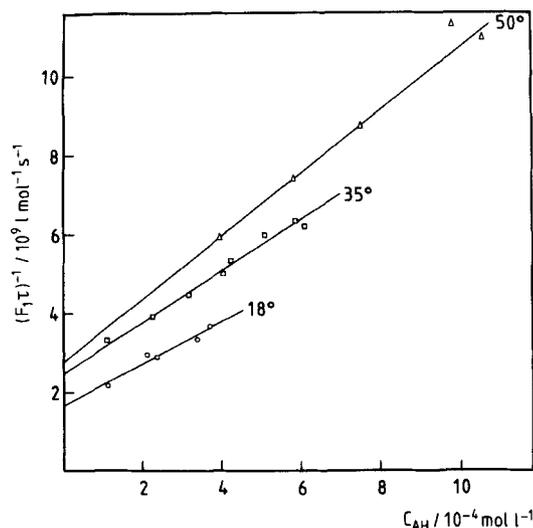


Fig. 4

Temperature dependence of the relaxation time for proton transfers between MNP and Acridine Orange. Plot of the rate expression Eq. (7) vs. equilibrium concentration of MNP

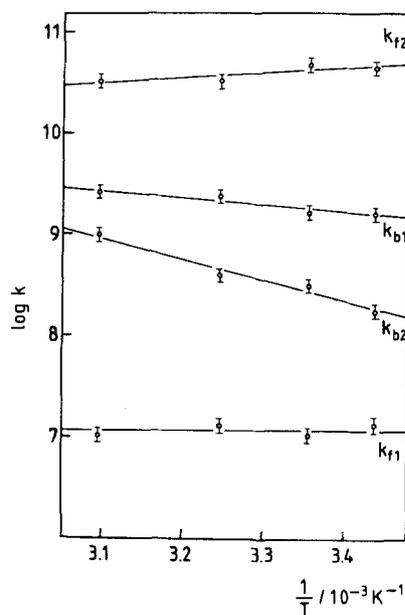


Fig. 5

Arrhenius plot of the rate constants in the Acridine Orange-MNP system

The errors are standard errors of the parameters of the best straight lines. The activation enthalpy  $\Delta\bar{H}_1^\ddagger = E_a - RT = 10.4/\text{kJ mol}^{-1}$  calculated from this result is within the confidence limits equal to the negative value of the corresponding reaction enthalpy  $-\Delta H_1 = 14.5/\text{kJ mol}^{-1}$ , and  $\Delta\bar{H}_2^\ddagger = 37.7/\text{kJ mol}^{-1}$  is even smaller than the negative of the reaction enthalpy  $-\Delta H_2 = 50.4/\text{kJ mol}^{-1}$  [8]. Thus the forward rate constant  $k_{f1}$  is practically independent of temperature, whereas  $k_{f2}$  decreases with increasing temperature, as shown in Fig. 5. The activation parameters are listed in Table 3.

The dependence of the rates on the ionic strength  $I$  was investigated at 25°C by varying the amount of the inert salt *TBHP* (Tables 1 and 2, Fig. 6). Rate constants were calculated as before using the equilibrium constants  $K_1$  and  $K_2$  measured independently at the same ionic strength.

The dependence of a diffusion controlled reaction rate on the viscosity and the ionic strength can be expressed in terms of the rate constant  $k_D^0$  at zero ionic strength, the viscosities of the solvent  $\eta_0$  and solutions  $\eta$ , and the activity term  $\Pi_f$  [10]

Table III  
Activation parameters and thermodynamic data for the reactions (1) and (2) of 3-Methyl-4-nitrophenol *MNP*

	$\text{AH} + \text{B} \rightleftharpoons \text{A}^- + \text{HB}^+$	$2 \text{AH} + \text{B} \rightleftharpoons \text{AHA}^- + \text{HB}^+$
$\Delta\bar{H}^\ddagger$	10.4	37.7
$\Delta\bar{H}^\ddagger$	-4.1	-12.7
$-\Delta H$	14.5	50.4
$\Delta\bar{S}^\ddagger$	-25	+51
$\Delta\bar{S}^\ddagger$	-115	-76
$-\Delta S$	90	127

Dimensions:  $\Delta H$ ,  $\text{kJ mol}^{-1}$ ;  $\Delta S$ ,  $\text{JK}^{-1} \text{mol}^{-1}$ .

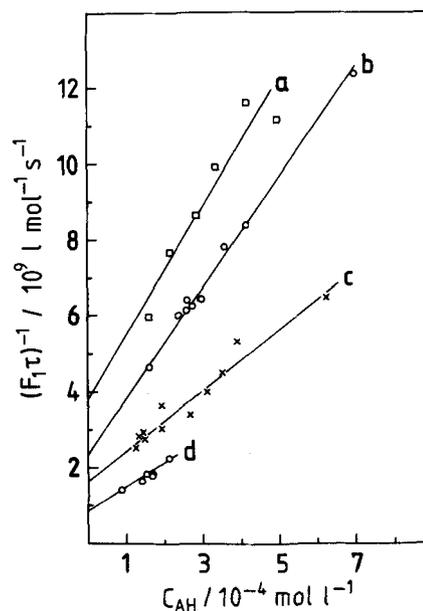


Fig. 6

Ionic strength dependence of the relaxation time for proton transfers between MNP and Acridine Orange at 25°C. Plot of the rate expression Eq. (7) vs. the equilibrium concentration of MNP at ionic strength  $I$ : a) 0.025 M; b) 0.05 M; c) 0.1 M; d) 0.2 M

$$k_D = \frac{\eta_0}{\eta} \cdot k_D^0 \cdot \Pi_f, \quad \Pi_f = \frac{f^+ f^-}{f^*} \quad (8)$$

where  $f^+$ ,  $f^-$ , and  $f^*$  denote the ionic activity coefficients of the cation, the anion and the activated complex respectively. By fitting the experimental rate constants  $k_{b1}$  to Eq. (8) using the Debye Hückel formula

$$-\log f = \frac{A \cdot \sqrt{I}}{1 + B \cdot r \cdot \sqrt{I}}$$

for the ionic activity coefficients  $f^+$  and  $f^-$ , and taking  $f^* = 1$  a value  $k_D^0 = k_{b1}^{0, \text{MNP}} = 1.8 \cdot 10^{10} \text{ M}^{-1} \text{ s}^{-1}$  and the ionic radius  $r = 0.43 \text{ nm}$  were calculated for the *MNP*/acridine orange system. The same calculation results in  $k_{b2}^{0, \text{MNP}} = 1.8 \cdot 10^9 \text{ M}^{-1} \text{ s}^{-1}$  and  $r = 0.89 \text{ nm}$  for the reaction of  $\text{HB}^+$  with the homoconjugate anion of *MNP*. The forward rates are independent of the ionic strength, as is expected for reactions between neutral molecules (Fig. 7).

In all calculations the ionic strength was assumed to be constant, and completely determined by the concentration of the inert salt *TBHP*. No activity terms need to be included in the kinetic Eqs. (6) and (9), because in no case the concentration of the reacting ions exceeded  $3 \cdot 10^{-5} \text{ M}$ , whereas the smallest concentration of *TBHP* was  $2.5 \cdot 10^{-2} \text{ M}$ .

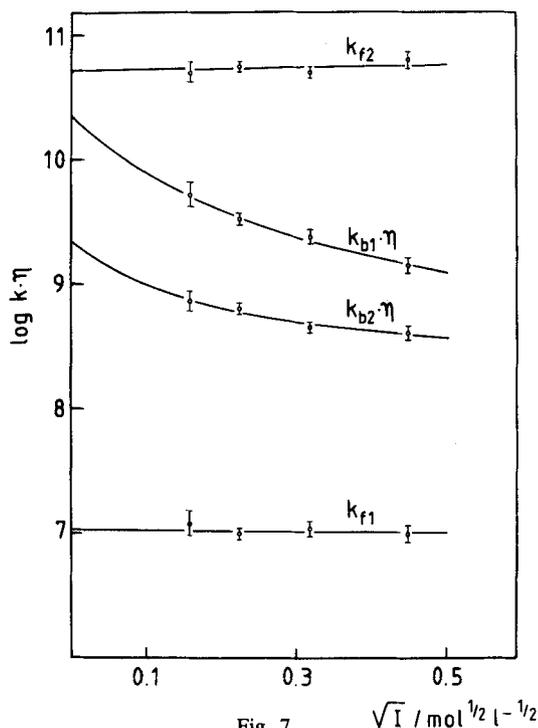
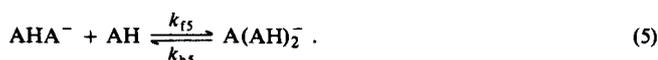


Fig. 7  $\sqrt{I} / \text{mol}^{1/2} \text{l}^{-1/2}$

Dependence of the apparent rate constants  $k_{f1}$  and  $k_{f2}$ , and of the rate constants  $k_{b1}$  and  $k_{b2}$ , corrected for the viscosity dependence, on the ionic strength  $I$ . The curves through  $k_{b1}\eta$  and  $k_{b2}\eta$  are the best fits of Eq. (8)

### 3.2. Protonation of Acridine Orange by Weakly Acidic Phenols

A  $10^2 - 10^3$  - fold excess of the weak acids *CMP* and *DMP* is necessary to protonate the acridine orange base to a measurable extent. Under these conditions only reactions (2) and (3) are observable [8]. As with the nitrophenols, only one chemical relaxation with a time constant in the  $10 - 50 \mu\text{sec}$  range was found (Table 4). This was attributed to the parallel reactions (2) and (3), which are coupled by the hydrogen bond association of the homoconjugate anion



If it is assumed that  $k_{f5} \geq 10^7 \text{ M}^{-1} \text{ s}^{-1}$  holds, the relaxation time of (5) is outside the timescale of our measurement for all phenol concentrations used. Thus the reciprocal relaxation time  $1/\tau$  for reactions (2) and (3) is given by

$$1/\tau = k_0 F_2, \quad (9)$$

with

$$F_2 = K_2 \left[ c_{\text{AH}}^2 + \frac{c_{\text{AH}} c_{\text{B}} (6 c_{\text{AH}} + 4 K_5^{-1})}{c_{\text{AH}} + c_{\text{AHA}^-} + K_5^{-1}} \right] + c_{\text{AHA}^-} + \frac{c_{\text{HB}} + (3 c_{\text{AHA}^-} + K_5^{-1})}{c_{\text{AH}} + c_{\text{AHA}^-} + K_5^{-1}}$$

accounting for the rapid pre-equilibrium. The observed rate constant  $k_0$  is defined by

$$k_0 = k_{b2} + k_{b3} \cdot K_5 \cdot c_{\text{AH}},$$

which allows the determination of  $k_{b2}$  and  $k_{b3}$  from intercept and slope of linear plots of Eq. (10)

$$(\tau F)^{-1} = k_{b2} + k_{b3} \cdot K_5 \cdot c_{\text{AH}} \quad (10)$$

shown in Fig. 8.

Table IV  
Reactions of Acridine Orange with Methylphenols at 25°C, ionic strength  $I = 0.1 \text{ M}$

$10^2 c_{\text{AH}}^0$ <sup>a)</sup> M	$10^5 c_{\text{B}}^0$ <sup>b)</sup> M	$\tau$ $\mu\text{sec}$
4-Cl-3,5-dimethylphenol		
0.983	35.2	45.3
2.45	2.08	42.6
1.36	24.9	34.5
2.61	4.46	25.9
2.20	12.4	22.0
1.77	23.3	21.5
2.60	17.8	17.9
4.03	3.00	17.3
3.19	10.8	13.8
3.69	8.63	13.2
3.70	12.0	12.7
4.56	1.62	10.9
3,5-Dimethylphenol		
8.27	2.11	58.6
9.92	2.68	38.2
12.4	1.91	30.5
7.94	13.1	25.9
12.2	4.16	23.3
9.71	9.77	22.2
6.86	28.2	22.2
8.50	18.4	21.5
11.1	7.44	21.0
10.2	13.5	19.1
10.7	11.6	18.9
12.8	7.33	17.3
9.45	25.4	14.8
15.4	6.26	13.1

<sup>a)</sup> Total concentration of phenol.

<sup>b)</sup> Total concentration of Acridine Orange.

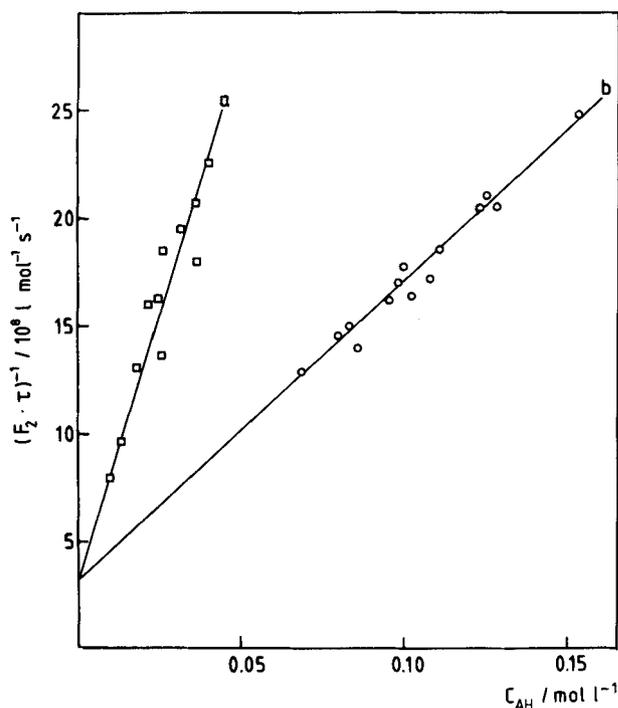


Fig. 8

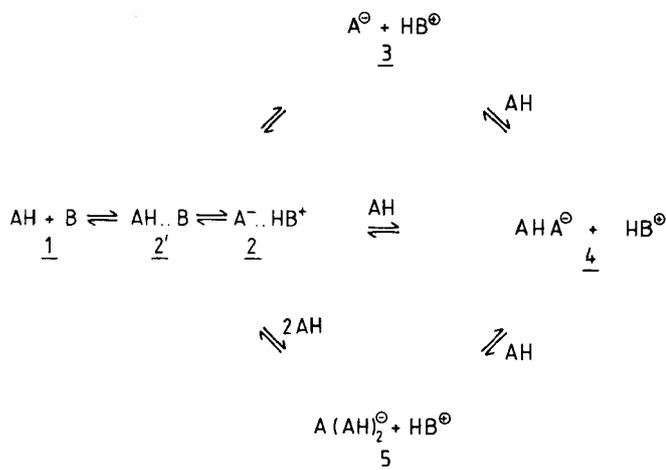
Dependence of the relaxation time of proton transfers between Methylphenols and Acridine Orange on the phenol concentration. Plot of the rate expression Eq. (10) vs. the equilibrium concentration of a) *CMP* and b) *DMP*





$K_{23} = \frac{C_{A^-} \cdot C_{HB^+}}{C_{A^- \cdots HB^+}} = 1.3 \cdot 10^{-3}$  M. Because of the hydrogen bond interaction in the ion pair  $K_{23}$  is significantly smaller than the purely electrostatic value  $2.8 \cdot 10^{-2}$  M calculated with the Fuoss-Eigen equation [10].

The full mechanism of proton transfer may be interpreted in the framework of an extended Eigen scheme including the different hydrogen bonded complexes (Scheme II).



The original Eigen mechanism, which is valid for proton transfer in protic solvents, consists of the elementary steps  $1 \rightleftharpoons 2' \rightleftharpoons 2 \rightleftharpoons 3$ . In benzonitrile the formation of hydrogen bonded complexes by direct reaction of the intermediate ion pair with acids in  $2 \rightleftharpoons 4$  and  $2 \rightleftharpoons 5$  makes an important contribution to the overall rate. At phenol concentrations larger than  $10^{-2}$  M only the latter steps are observed. Under these conditions the free anion is not even an intermediate in the acid base reaction.

According to Eigen only the proton transfer within the complex  $2' \rightleftharpoons 2$  depends on the difference of the pK-values of acid and protonated base  $\Delta pK$ . The same argument has been used by Marcus in the treatment of proton transfer reactions, in which the transfer step and not diffusion is rate limiting [25].

Marcus obtained a relation between the free energy of activation and the free energy of reaction in terms of the intrinsic energy barrier of height  $\lambda/4$  which must be overcome in the proton transfer step  $2 \rightleftharpoons 2'$  on the one hand and the work  $W$  required to bring the reactants (or products) together to form the reaction complex on the other hand. The intrinsic barrier  $\lambda/4$  is associated with the need for electron redistribution and alterations of bond length and bond angles during the course of the proton transfer. The work term  $W$  also includes the entropy of localization, the free energy of solvation, and the energies due to steric and statistical factors which are required in order to form the reaction complex. When the dependence of proton transfer rates on the structure of the reactants is interpreted along these lines, the assumption is generally made that only  $\lambda$  depends on  $\Delta pK$ , the work term  $W$  being treated as a constant [26, 27].

Our results provide for the first time direct experimental evidence related to these assumptions, and they also indicate possible limitations. As is shown in the previous sections the stability constant of the ion pair  $K_{12}$  parallels almost exactly the

acidity of the phenol, whereas the rates and the equilibrium constants of the dissociation of the ion pair to the free ions  $2 \rightleftharpoons 3$  and of the formation of the hydrogen bonded anion  $2 \rightleftharpoons 4$  are constant. This allows one to attribute the dependence of the overall forward rate constants  $k_{f1}$  and  $k_{f2}$  and of  $K_{13}$  and  $K_{14}$  to the change of  $K_{12}$  with the acidity of the phenol. This was predicted by Eigen, and is in agreement with Marcus treatment since the barrier height  $\lambda/4$  depends on  $\Delta G^0$ , the difference in free energies of the proton in  $2'$  and  $2$ . It must be noted, however, that the reaction  $2 \rightarrow 4$  includes the solvation of the anion by a phenol molecule, whose proton donating ability decreases to the same extent as the basicity of the anion increases in a series of related substances. If, instead, a proton donor with constant properties would solvate the anions, no constancy of either  $k_{24}$  or  $K_{24}$  within the series could be expected.

Also, the second step of anion solvation shows a significantly different structure reactivity relationship. In  $2 \rightleftharpoons 5$  the loose hydrogen bonded complexes  $A(AH)_2^-$  ( $\Delta H_{45} \sim 0$  kJ mol $^{-1}$ ) react with the protonated acridine orange  $HB^+$  at almost a diffusion controlled rate. The observed rate constants  $k_{b3}$  may be interpreted as  $k_{52}$  in analogy to the other ion recombinations. Since  $k_{52}$  does not depend on the acidity of the phenol, whereas  $K_{45}$  does, the forward rate constants  $k_{25}$  are different for different phenols in contrast to the assumed constancy of  $k_{24}$ . Certainly, an additional hydrogen bonded intermediate is involved in  $2 \rightleftharpoons 5$ ; the detailed formulation of this pathway does not change the argument, however. Since the separate elementary steps of anion solvation by hydrogen bonding depend in a different way on the properties of the proton donor constancy of solvation terms in the energetics of proton transfer in a protic solvent may occur only by fortuitous cancellation of different effects.

The authors are indebted to Prof. H. W. Zimmermann for his constant encouragement, and to Prof. C. F. Bernasconi, Prof. M. M. Kreevoy and Dr. H. H. Limbach for helpful discussions. Acknowledgement is made to the Deutsche Forschungsgemeinschaft, the Fonds der Chemischen Industrie, and the Freiburger Wissenschaftliche Gesellschaft for financial support. R. S. gratefully acknowledges a stipend from the Studienstiftung des Deutschen Volkes. We are indebted to Mr. K. Krtischil for assistance with some of the calculations. The calculations were carried out at the Rechenzentrum of the University of Freiburg.

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(Eingegangen am 15. August 1983)

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## Absorption and Fluorescence Spectra, $pK_a$ Values, and Fluorescence Lifetimes of Monohydroxyflavones and Monomethoxyflavones

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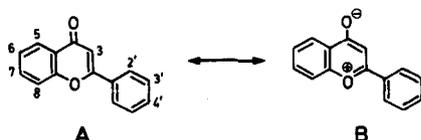
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### Fluorescence / Fluorescence lifetime / Ionization / Light, Absorption / Light, Emission

The absorption and fluorescence spectra of all isomeric hydroxyflavones and methoxyflavones, except for the photolabile 3-methoxyflavone, have been measured in organic solvents and in aqueous solutions of various acidity. With the exception of 5-hydroxyflavone, all are found to be fluorescent in methanol and water solution. On the other hand, there is virtually no fluorescence observed in cyclohexane solution, a fact that is interpreted in terms of high intersystem crossing rates and the El-Sayed selection rules. – Except for the 3-hydroxy and 7-hydroxy isomers, the anions of hydroxyflavones are non-fluorescent, whereas the protonated forms exhibit strong fluorescence emission. Methoxyflavones have higher fluorescence quantum yields than the corresponding hydroxyflavones and have fluorescence decay times ranging from 1.3 to 6.9 ns in methanol at room temperature. – The ground state  $pK_a$  values of the hydroxyflavones range from 7.8 to 9.8, except for the 5-hydroxy isomer (11.6). The  $pK_a$ 's governing the protonation step range from  $-1.22$  to  $-1.55$ , again with the exception of the 5-hydroxy isomer ( $-3.1$ ). The dissociation constants of the first excited singlet state were calculated with the help of the Förster-Weller equation. The results predict a reversal of the most basic and most acidic sites of the hydroxyflavones. In aqueous solutions, this should result in the formation of excited state tautomers which, however, could be detected only for the 3-hydroxy and 7-hydroxy isomers. Apparently, the lifetimes of the other isomers are too short to allow the establishment of excited state equilibria.

### Introduction

The flavones form one of the largest group of organic natural products [1–3]. Their occurrence is practically confined to the plant kingdom, being partially responsible for deep yellow colors of certain flowers. Among their biological activities [1, 2], flavones have been shown to act against blood cell adhesion and aggregation.



Scheme 1

Chemical structure of the flavone chromophore (A) and its dipolar resonance structure B

The structure and numbering of flavone is shown in Scheme 1. By analogy to the behavior of pyrones, flavone (A) can be formulated as a zwitterion (B), which contributes significantly to the flavone structure and can explain the relative basicity of the carbonyl groups.

Most natural flavones bear either hydroxy or methoxy groups or are linked to a sugar residue ("glycones"). Due to their importance, the absorption spectra have been studied in great detail [3, 5], since absorptiometry is one of the most suitable methods for the structure elucidation and identification of flavonoids [3, 5]. Interestingly, most of the data are confined to methanol as a solvent. The published spectra [6, 7] for the 6- and 8-hydroxy isomers do not agree with ours.

Except for our previous work on the pH-dependent fluorescence of flavonol [8], 7-hydroxyflavone [9] and 4'-hydroxy-